

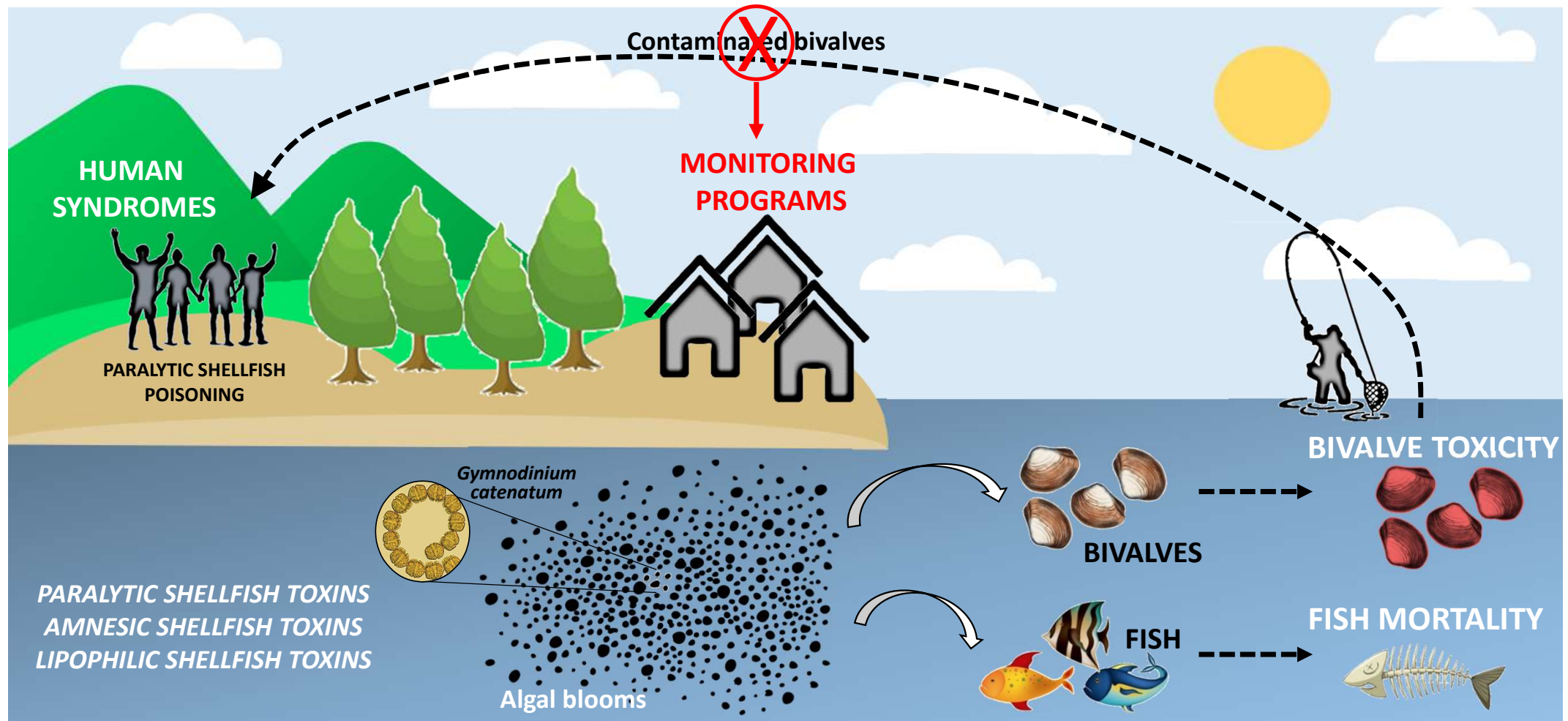
Deteção em tempo real de biotoxinas, será possível? Projectos LinguaTox e ImageTox

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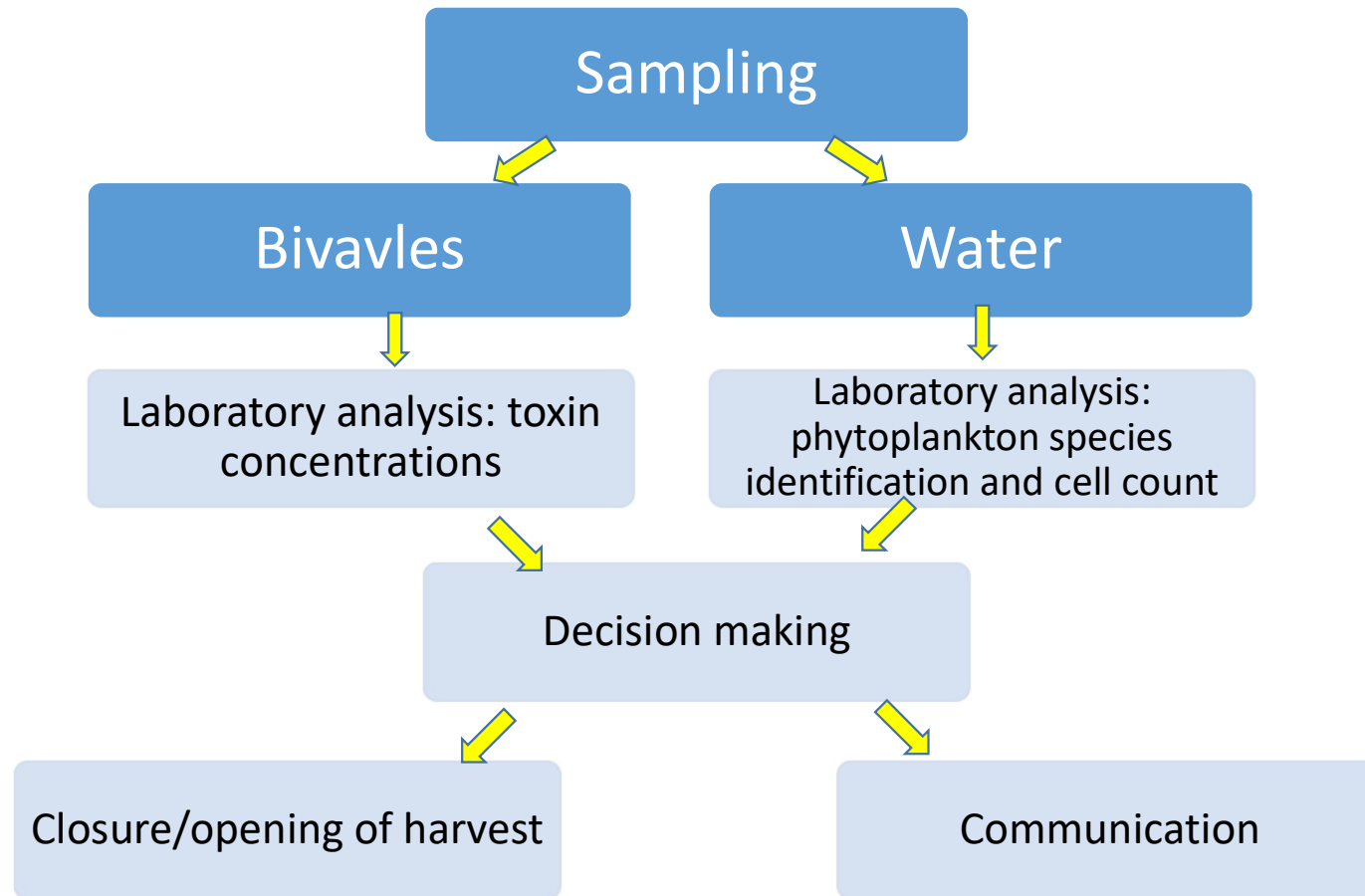
Seminário de divulgação de projetos da UNIVERSIDADE DE AVEIRO
apoiados pelo **Mar 2020** | 30 SETEMBRO 2022 | ÍLHAVO ECOMARE



Harmful algal blooms and bivalve toxicity



Toxin monitoring programs - National Reference Laboratories



Toxin analysis



Tissue removal



Tissue homogenization



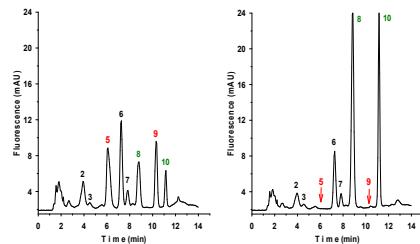
Toxin extraction



Sample preparation



Analysis by LC-FLD, LS-MS



Toxins' quantification and identification

Identification of toxic phytoplankton species e cell counting



Water column sampling

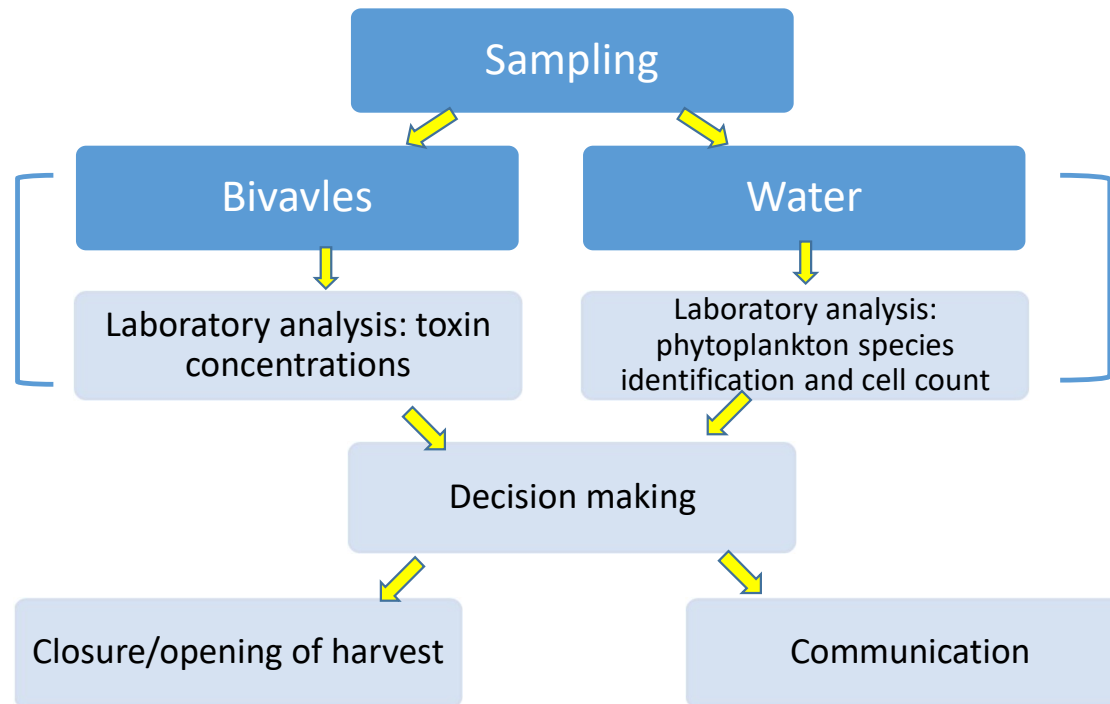


Sedimentation of fixed samples (Utermöhl method)



Identification and enumeration using inverted light microscopy

Toxin monitoring programs



promar
Programa Operacional Pesca 2007 - 2013

LínguaTox -
Língua bioelectrónica
para a detecção de
toxinas marinhas em
bivalves
2014-2015
Funding 279 175 €

mar
2020
PROGRAMA OPERACIONAL MAR 2020

ImageTox –
Detecção automática de
microalgas tóxicas
através de imagens
hiperespectrais
2018-2022
Funding 211 237 €

LinguaTox – Bioelectronic tongue for the detection of marine toxins in bivalves

Objective: development of the bioelectronic tongue based on chemical sensors and enzymatic biosensors for the rapid detection of Paralytic Shellfish Toxins (PSTs) detected on the Portuguese coast (decarbamoyl and N-sulfocarbamoyl).

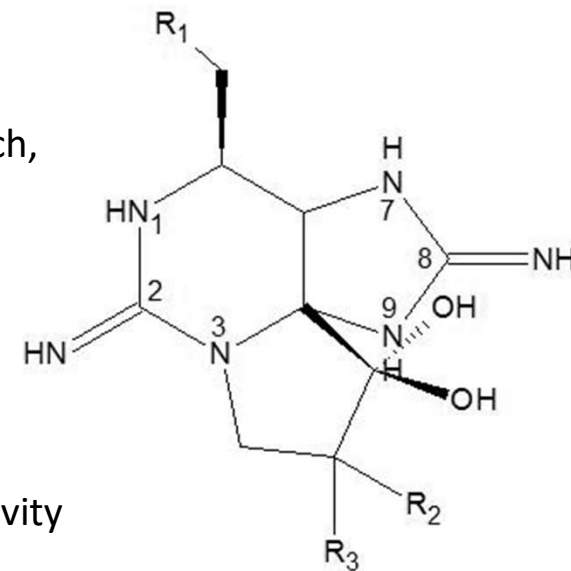
Why PSTs?

High toxicity and severity of symptoms:

- Poisoning symptoms: tingling, burning, numbness, drowsiness, incoherent speech, respiratory paralysis.
- Severity: respiratory support within 12 hours of exposure results in complete recovery; mortality 8.5 - 9.5 %.

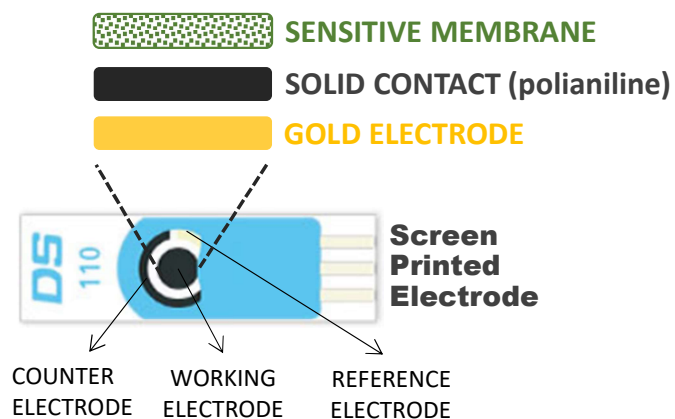
Challenge

- More than 50 PSTs have been identified
- PST profiles in bivalves depend on toxin-producing species and bivalve enzymatic activity
- Existing sensors and assays not applicable to N-sulfocarbamoyl PSTs' detection

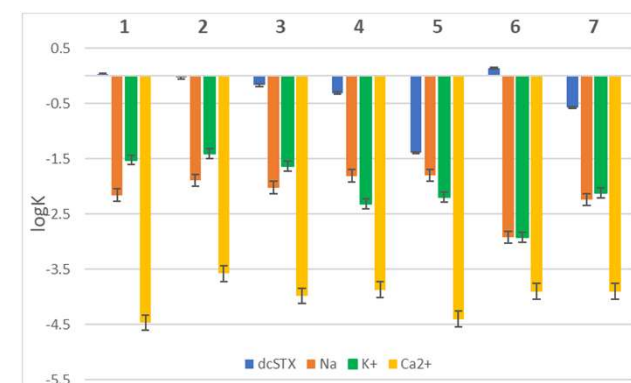
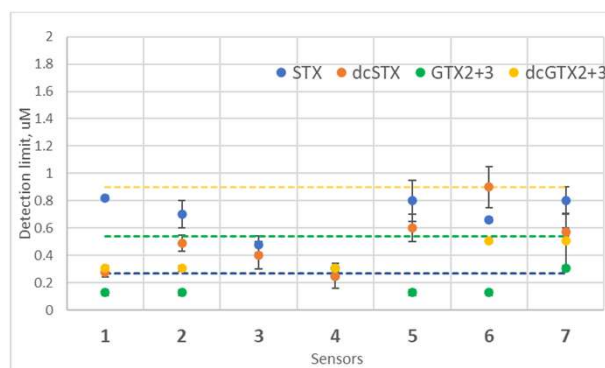
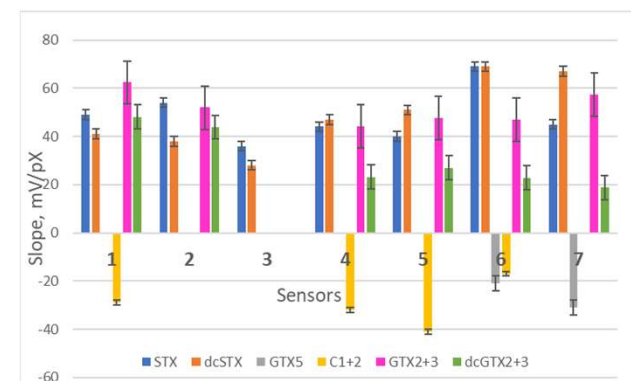


LinguaTox – chemical sensors' development

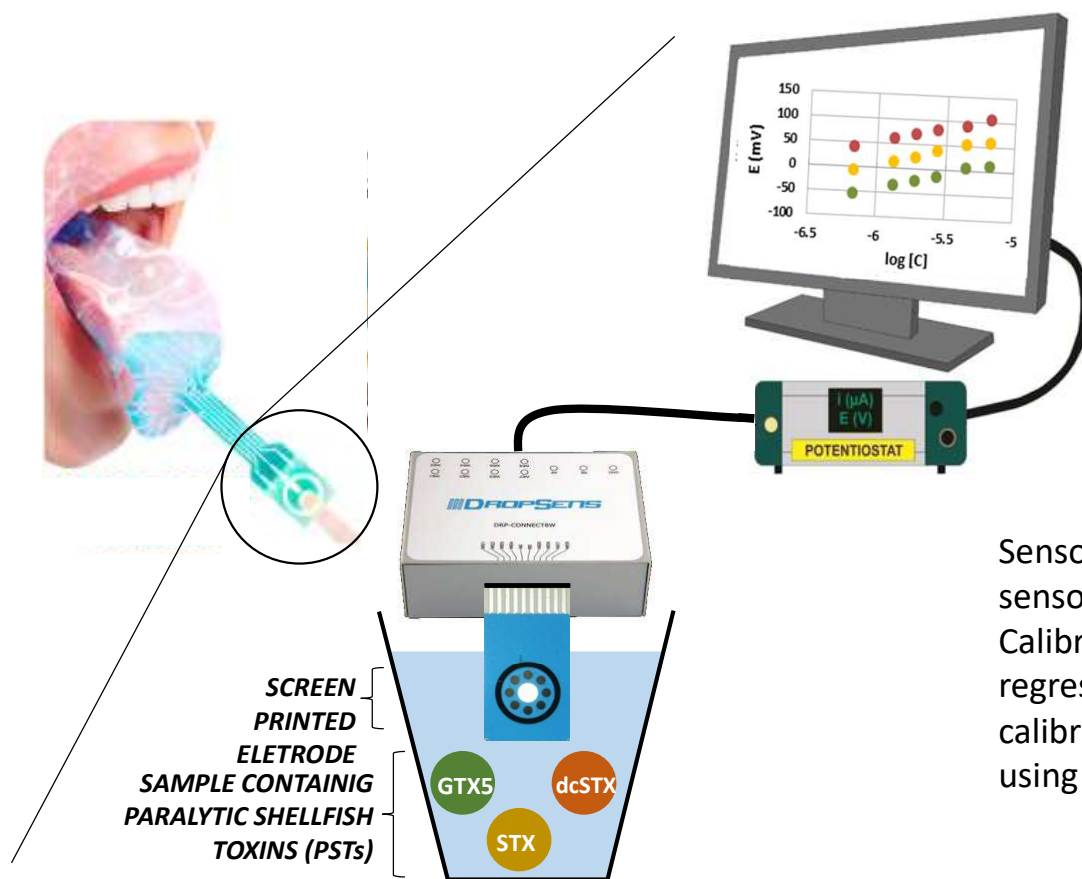
Potentiometric chemical sensors
with PVC membranes



High sensitivity and selectivity to
carbamoyl (STX, GTX2+3) and
decarbamoyl (dcSTX, dcGTX2+3)
Low sensitivity to N-
sulfocarbamoyl PSTs (GTX5, C1+2)

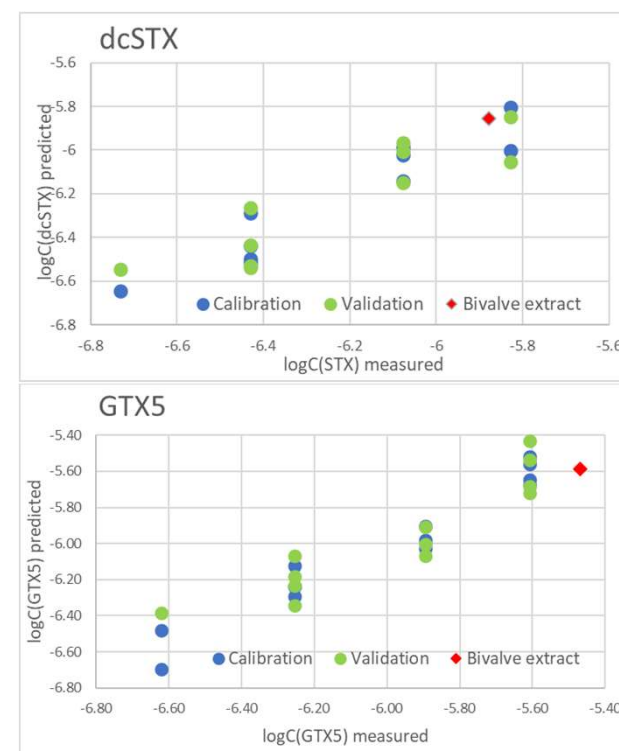


LinguaTox – electronic tongue development



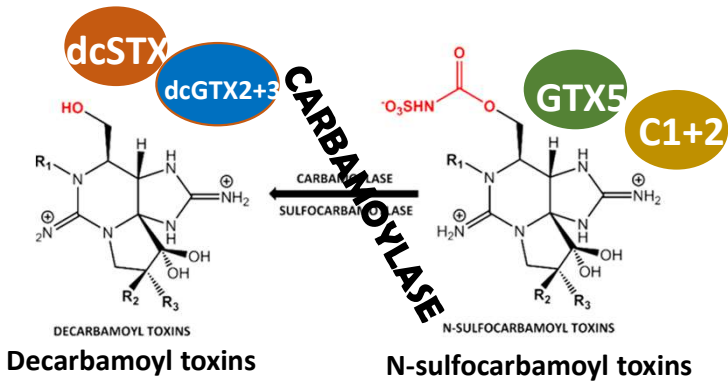
Sensor array – 6 sensors
Calibration – PLS-regression + calibration update using Joint Y-PLS

Quantification of STX, dcSTX and GTX5 in mussel extracts

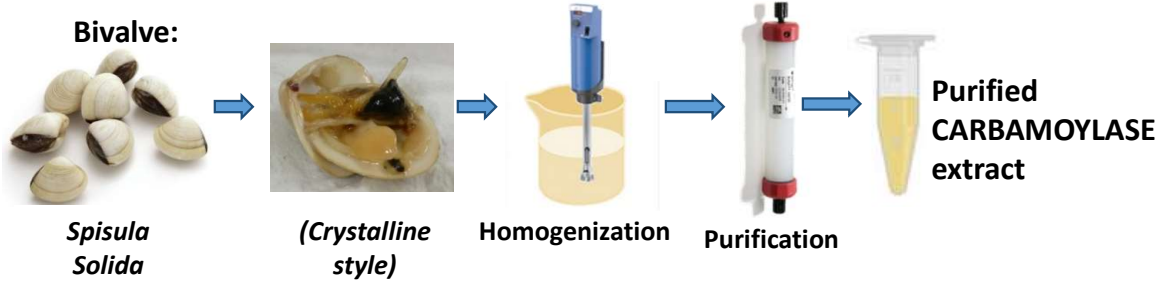


LinguaTox - Enzymatic biosensors for detection of the N-sulfocarbamoyl PSTs

PST transforming enzymes



Carbamoylase extraction



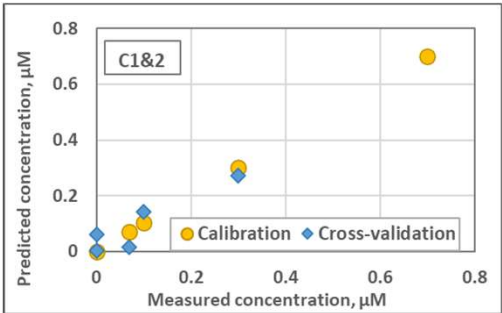
N-sulfocarbamoyl PSTs detection



Potentiometric detection of decarbamoyl PSTs



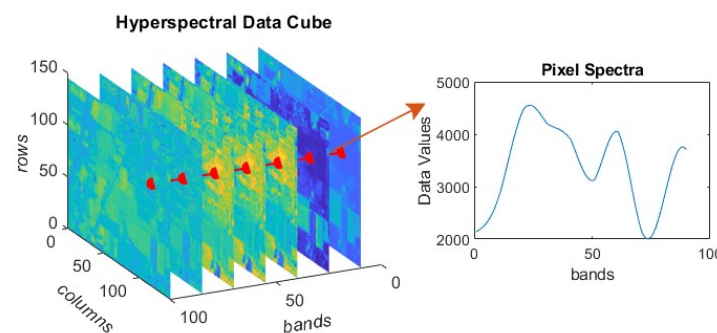
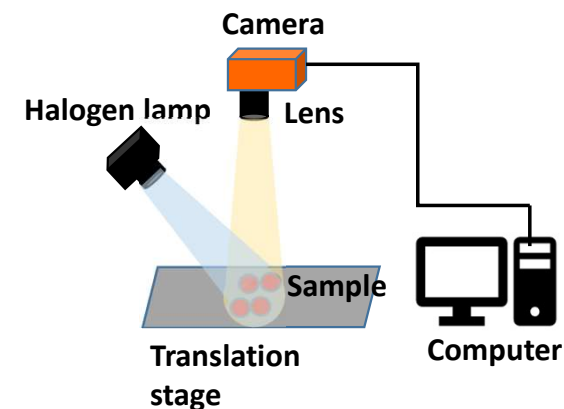
Impedimetric detection of enzymatic activity



ImageTox – Automatic detection of toxic microalgae using hyperspectral imaging

Objective: development an inexpensive, reagentless and easily automatable tool for the early detection and identification of microalgae species responsible for harmful algal blooms (HABs) based on the statistical analysis of the hyperspectral infrared images of phytoplankton.

Hyperspectral imaging



Spectral range: visible – far LWIR
(400 nm – 5.3 μm)

Applications:

Industry

- Food
- Pharmaceutical
- Plastic sorting
- Packaging
- Mineral exploration

Agriculture
Research

Operational environment:

- Airborne
- Field
- Plant
- Lab

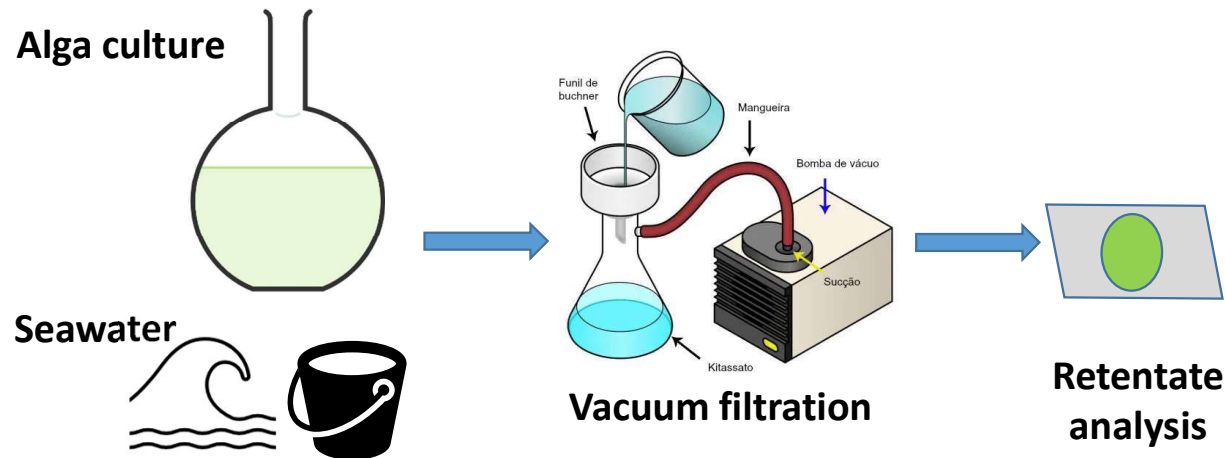
ImageTox - samples

Identification and quantification of toxic microalga in water samples in the presence of a mixed phytoplankton assemblage and its discrimination from nontoxic species with similar morphology using NIR hyperspectral imaging.

Comparison of NIR hyperspectral imaging with well-established spectroscopic techniques - NIR and FT-MIR spectroscopy.

Species selected for the study: *Gymnodinium*: *G. catenatum* and *G. nolleri*; *Pseudo-Nitzschias*: *P. australis* and *P. fraudulenta*; *Prorocentrum lima*.

Sample preparation



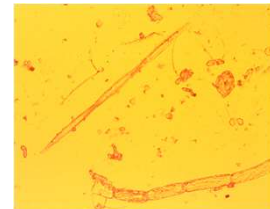
- Cultures of *G. catenatum* (toxic) and *G. nolleri* (nontoxic);
- Natural seawater collected at the Aveiro lagoon;
- Mixtures of seawater and algal cultures at cell concentrations 200, 500, 1000, 1500, 2000 and 5000 cells/L, selected to encompass the warning (500 cells/L) and closure (1500 cells/L) thresholds established in EU regulation for *G. catenatum*.



G. catenatum



G. nolleri



Seawater

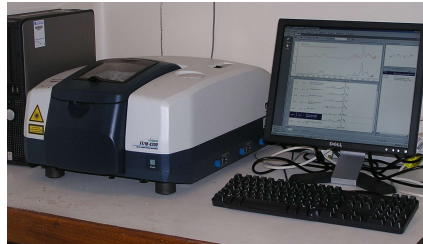
ImageTox – measurements and experimental set-up

NIR spectroscopy



Perkin Elmer Spectrum 100N
10000-4000 cm^{-1}

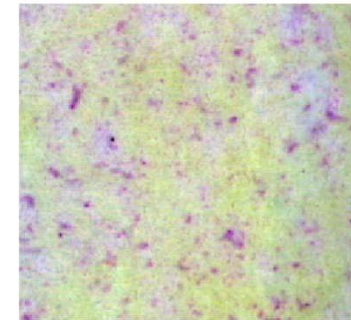
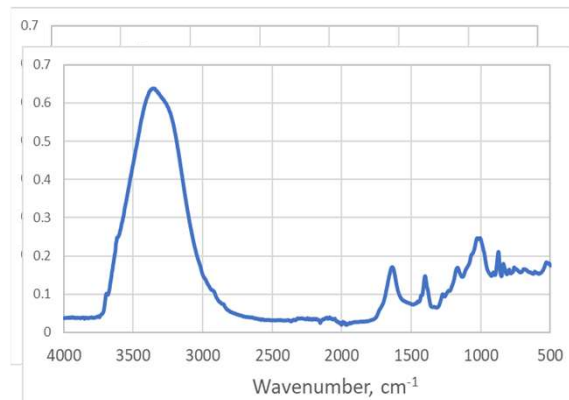
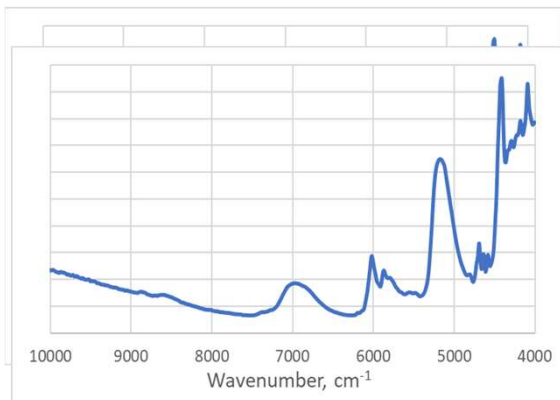
FT-MIR spectroscopy



Bruker Alpha Platinum with ATR accessory
4000-600 cm^{-1}

NIR HSI

LabSCANNER 40x20
FX17 spectral camera,
900 – 1700 nm
Macro lens OLES
MACRO (SPECIM, Ltd).
KL 1500 HAL Halogen;
Annular fiber optic
Darkfield ringlight
(SCHOTT AG)



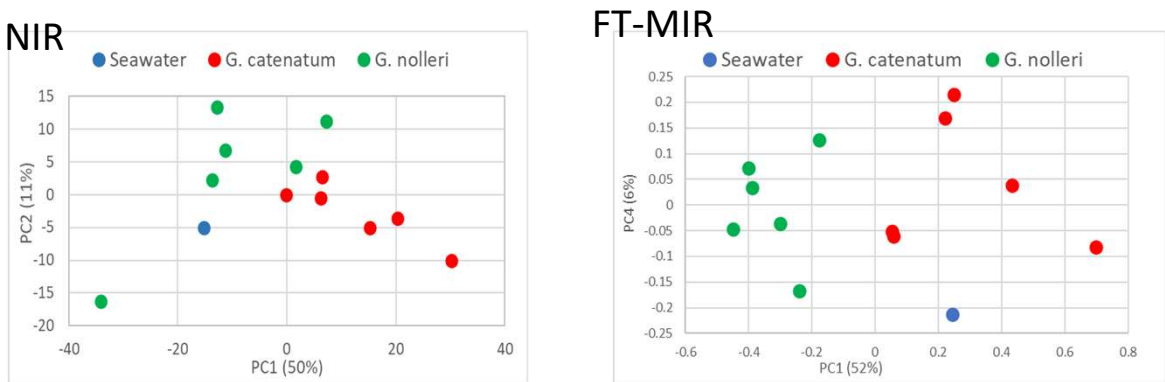
ImageTox - Data processing

	NIR spectra	FT-MIR spectra	NIR imaging
Data pre-processing	• Smoothing (Savitsky-Golay) • SNV normalization • Scaling	• Smoothing (Savitsky-Golay) • SNV normalization • Calculation of 2^d derivative • Scaling	• Smoothing in the spectral mode (Savitzky-Golay) • SNV normalization in the spectral mode • Endmember signature extraction • Abundance maps of endmember signature corresponding to <i>G. catenatum</i>
Data exploration		PCA	Abundance maps w.r.t. <i>G. catenatum</i> endmember signature
Classification		PLS-DA	NPLS-DA

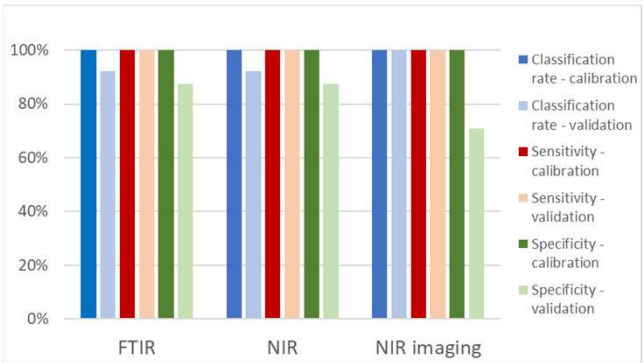
ImageTox

Case study *G.catenatum* and *G. nollery*

PCA score plot of seawater and its mixtures with cultures of two *Gymnodinium* species

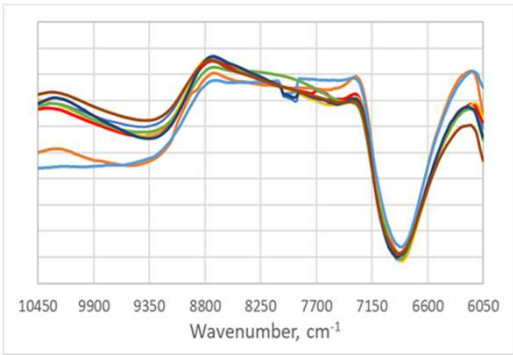


Classification results

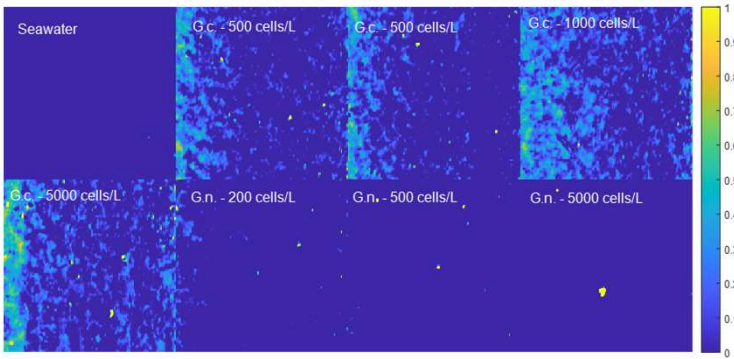


NIR Imaging

Endmembers signatures

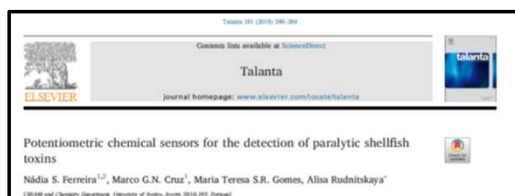


Abundance maps of endmember signature corresponding to *G. catenatum*



LinguaTox

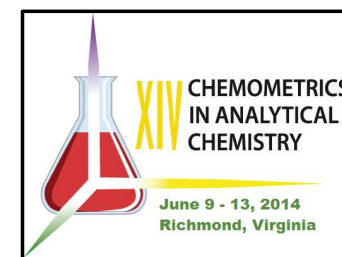
Articles:



Oral Presentations:



Poster:



ImageTox

Oral Presentations:



Poster:





Project team

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Thank you for your attention!

